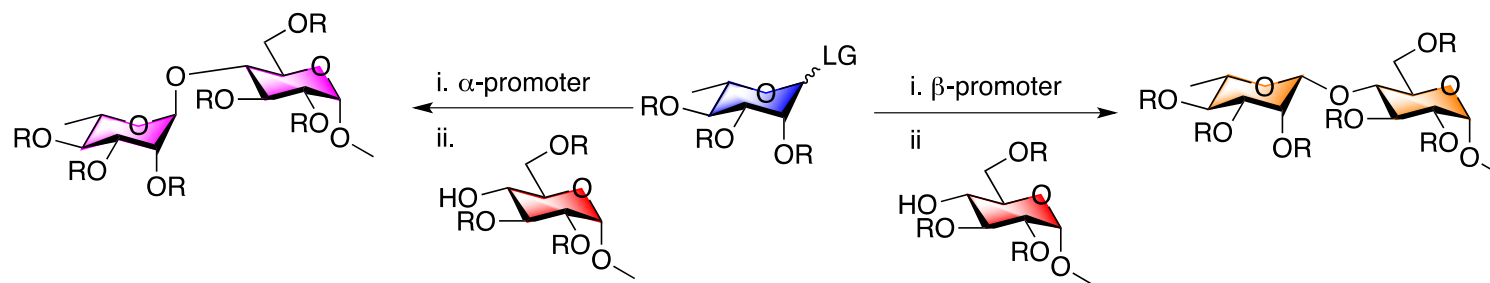
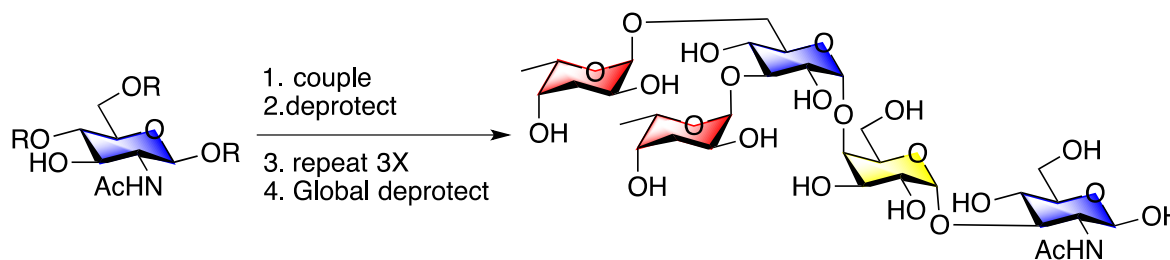


# Common Fund Research in the Bennett Group: Reagent Controlled Glycosylation (U01GM120414-01)

*Developing Chemical Promoters that Permit Absolute Control Over the Stereochemical Outcome of Glycosylation Reactions:*



*These Promoters will Make Oligosaccharide Construction Similar to Peptide Synthesis:*

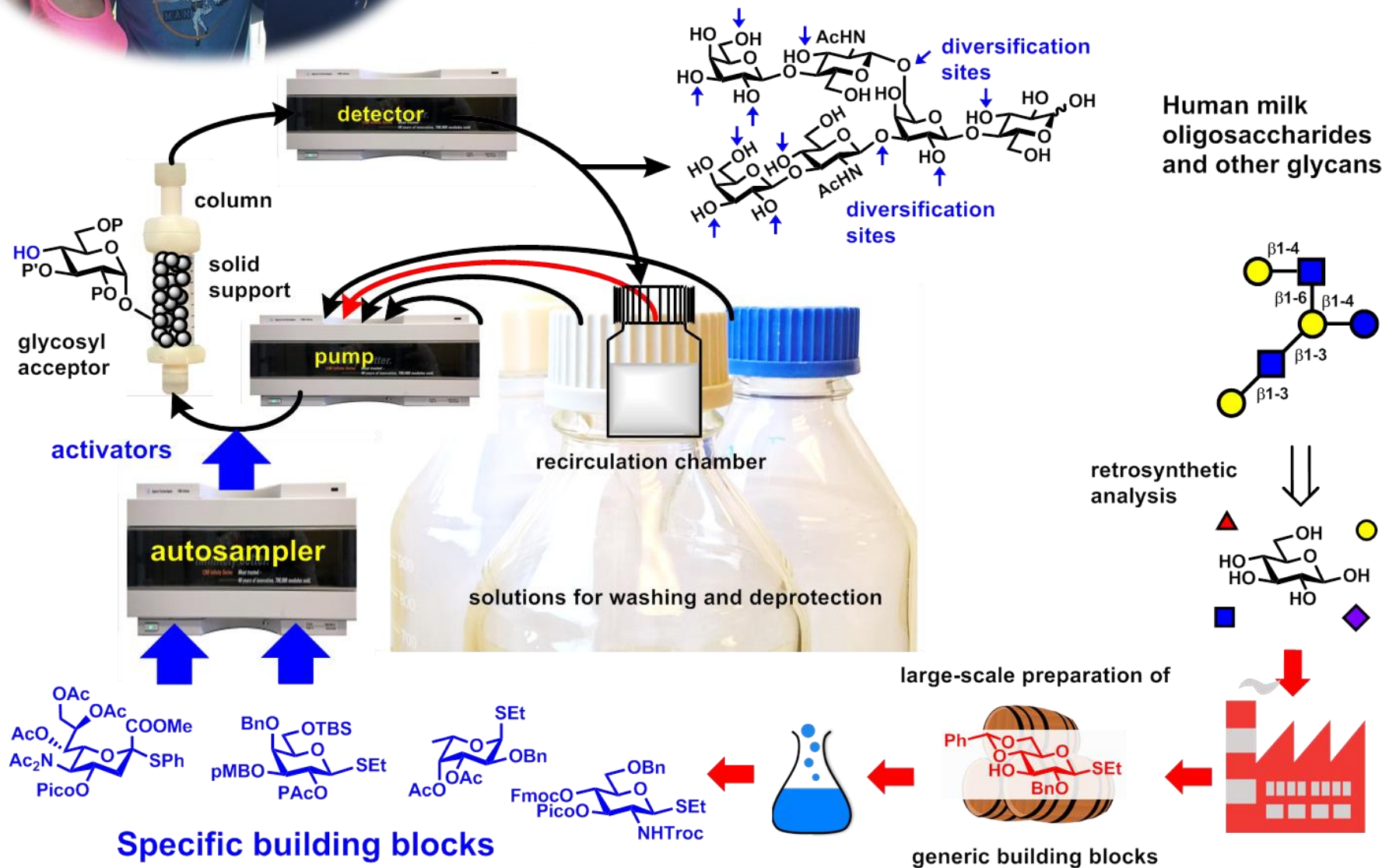


*Rapid Construction of Oligosaccharides for Analytical Standards and Therapeutic Development!*

# Refinement and implementation of the automated oligosaccharide synthesizer (U01GM120673, 2016-)



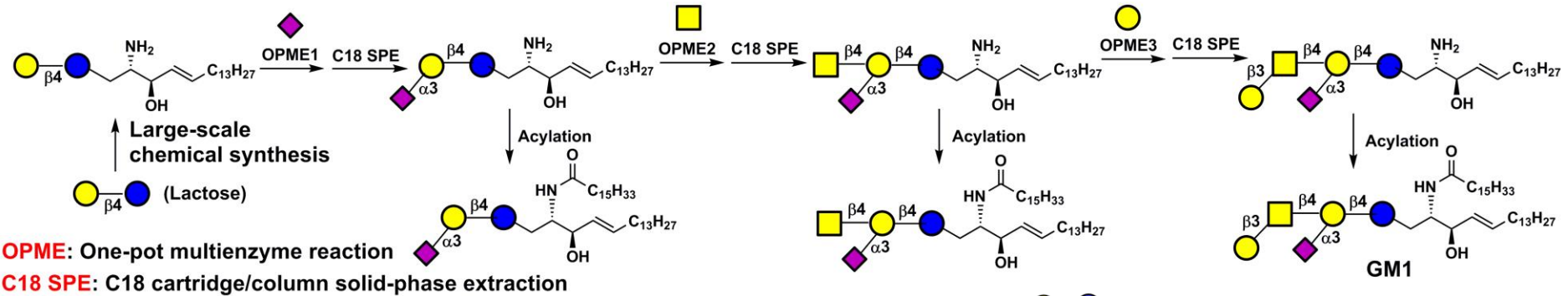
Alexei V. Demchenko, Keith J. Stine, University of Missouri - St. Louis  
 & Cristina De Meo, Southern Illinois University, Edwardsville



# Facile chemoenzymatic synthesis and purification of glycolipids

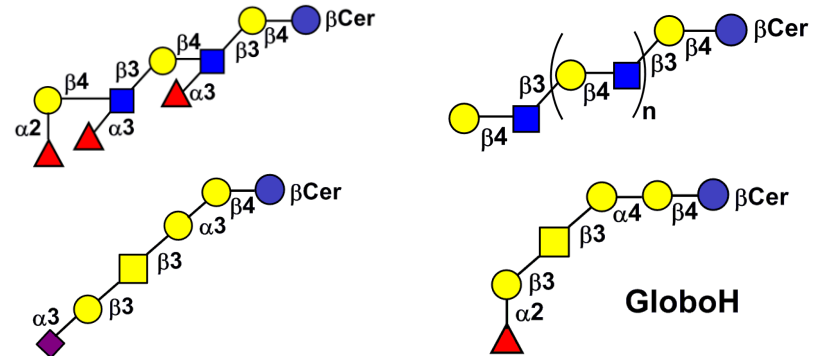
## NIH Common Fund Glyco-science Program (U01GM120419)

Xi Chen, U. of California-Davis, [xiichen@ucdavis.edu](mailto:xiichen@ucdavis.edu), <http://chengglyco.faculty.ucdavis.edu/>  
 Peng G. Wang, Georgia State U., [pwang11@gsu.edu](mailto:pwang11@gsu.edu), <http://lithium.gsu.edu/faculty/PWang/>



### Glycosphingolipids

- ganglio-series
- (neo)lacto-series, fucosylated and sialylated
- (iso)globo-series



**Goal:** To allow non-specialists to synthesize, functionalize, purify, and study glycosphingolipids

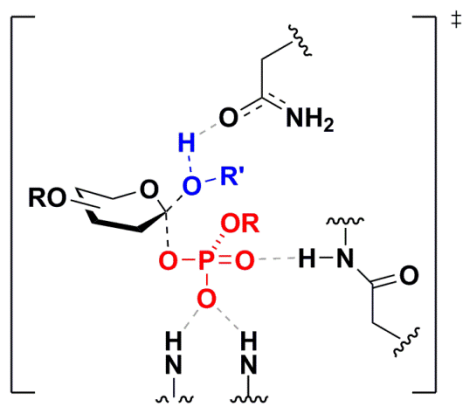
- Identify stable storage conditions for enzymes and reagents
- Assemble OPME enzyme and reagent kits
- Optimize reaction and purification conditions
- Establish protocols for OPME synthesis and C18 cartridge/column purification
- Cross-validation
- For more information, see <http://chengglyco.faculty.ucdavis.edu/glycosphingolipids/>

	<b>Glc</b>
	<b>Gal</b>
	<b>GlcNAc</b>
	<b>GalNAc</b>
	<b>Fuc</b>
	<b>Sia</b>

# Jacobsen Group

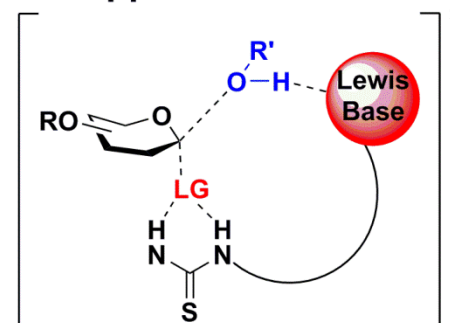
Developing Catalysts for Selective Glycosylation

Nature's solution:



- Selectivity through stereospecificity

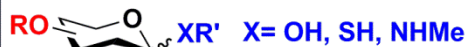
Our approach:



- Mild reaction conditions through dual-activation strategy
- Catalyst-promoted stereospecificity

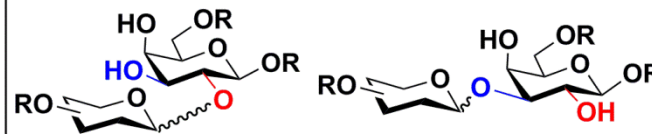
New strategy enables:

- Broad nucleophile, electrophile scope

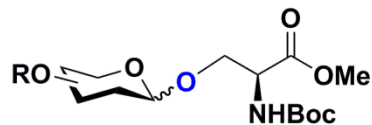


R = OBn, Me, OAc, NHAc, N<sub>3</sub>

- Catalyst control of multiple reactive sites

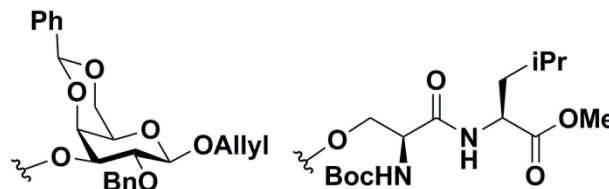


- High anomeric selectivity, efficient reactivity



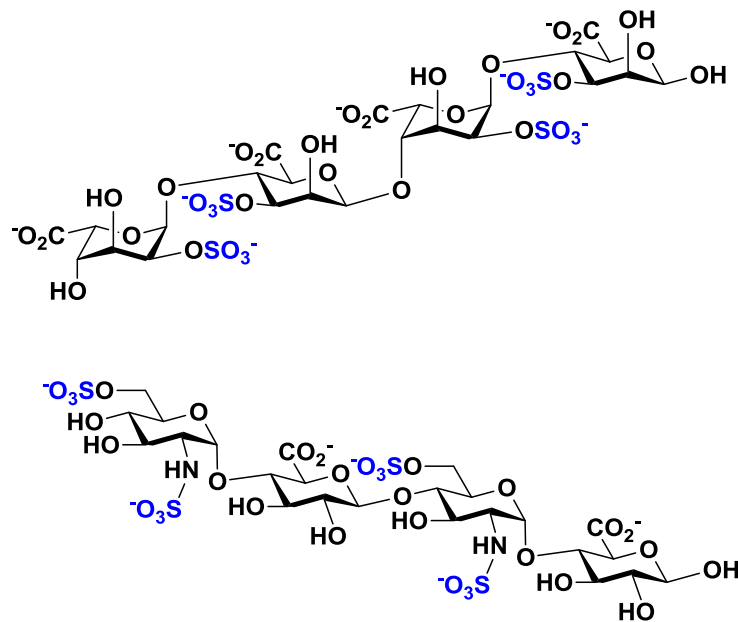
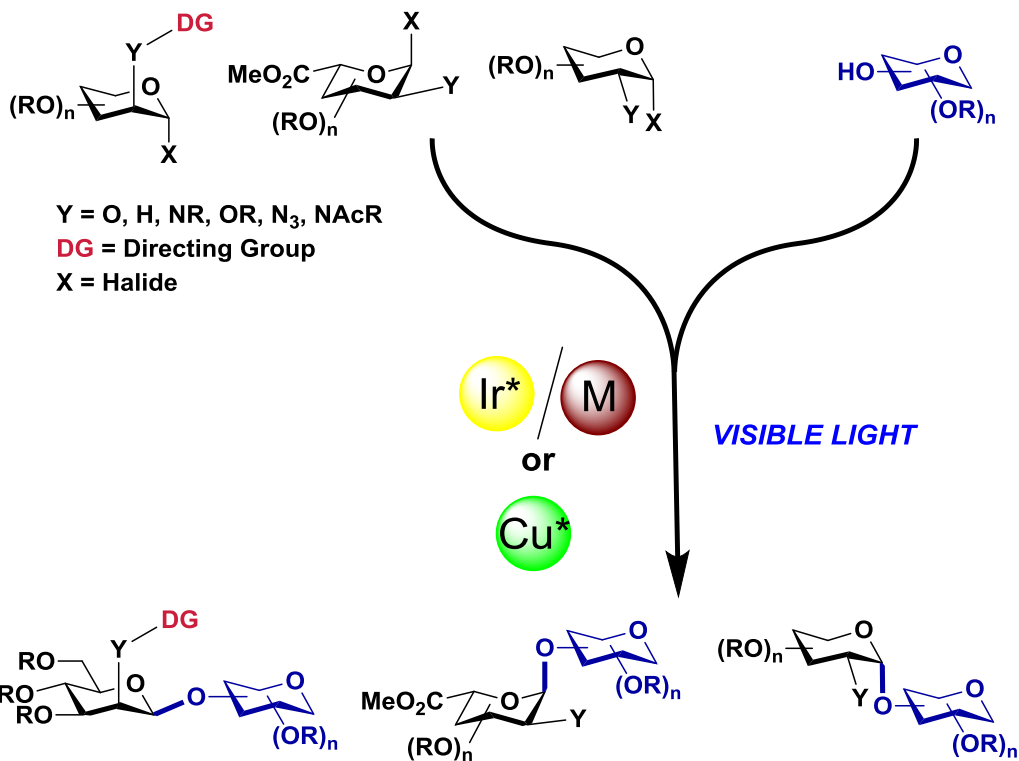
>20:1  $\beta$ : $\alpha$ , < 4 hours

- High functional group compatibility



# NIH Common Fund Research in the Nguyen Group: Stereoselective 1,2-Cis Glycosylation

Developing predictable and stereoselective 1,2-cis glycosylation reactions via either dual catalytic photoredox catalysis or photoinduced copper catalysis



*Rapid and stereoselective synthesis of bioactive oligosaccharides for analytical standards and therapeutic applications*

NIH-U01 GM120293

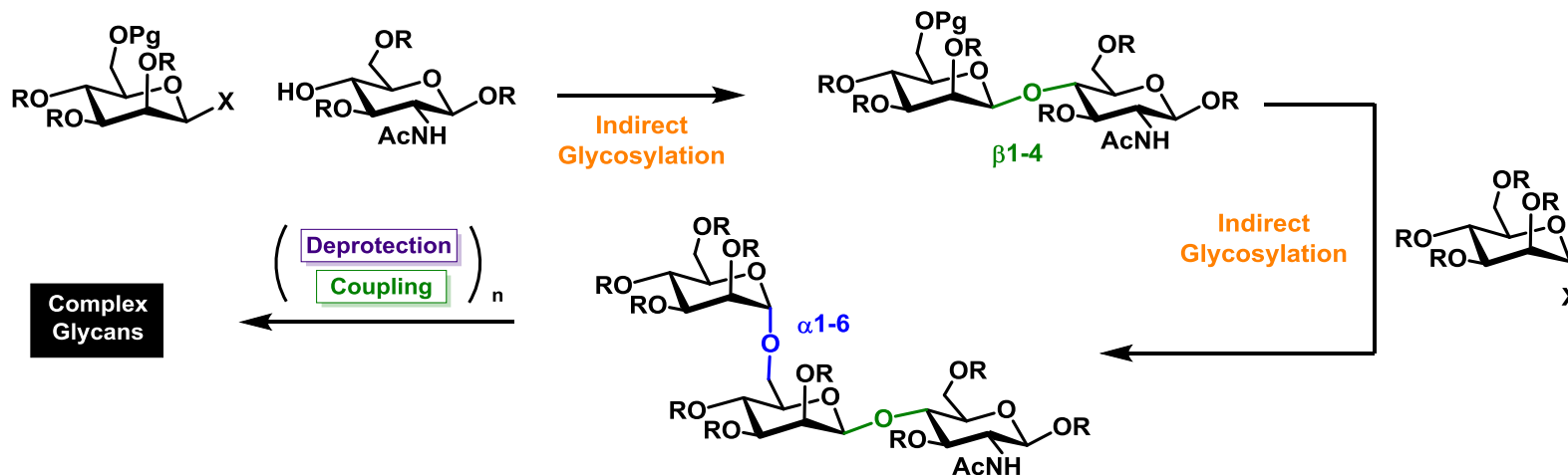
<https://commonfund.nih.gov/Glycoscience>

Hien M. Nguyen (hien-nguyen@uiowa.edu)  
University of Iowa

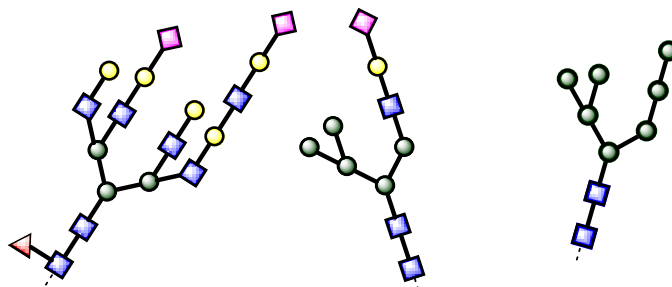
<https://nguyenresearchgroup.lab.uiowa.edu/>

# NIH Common Fund Research in the Brichacek Group: Novel Glycosylation Mechanisms

*Indirect Glycosylation Methods to Facilitate More Efficient and Selective Couplings*

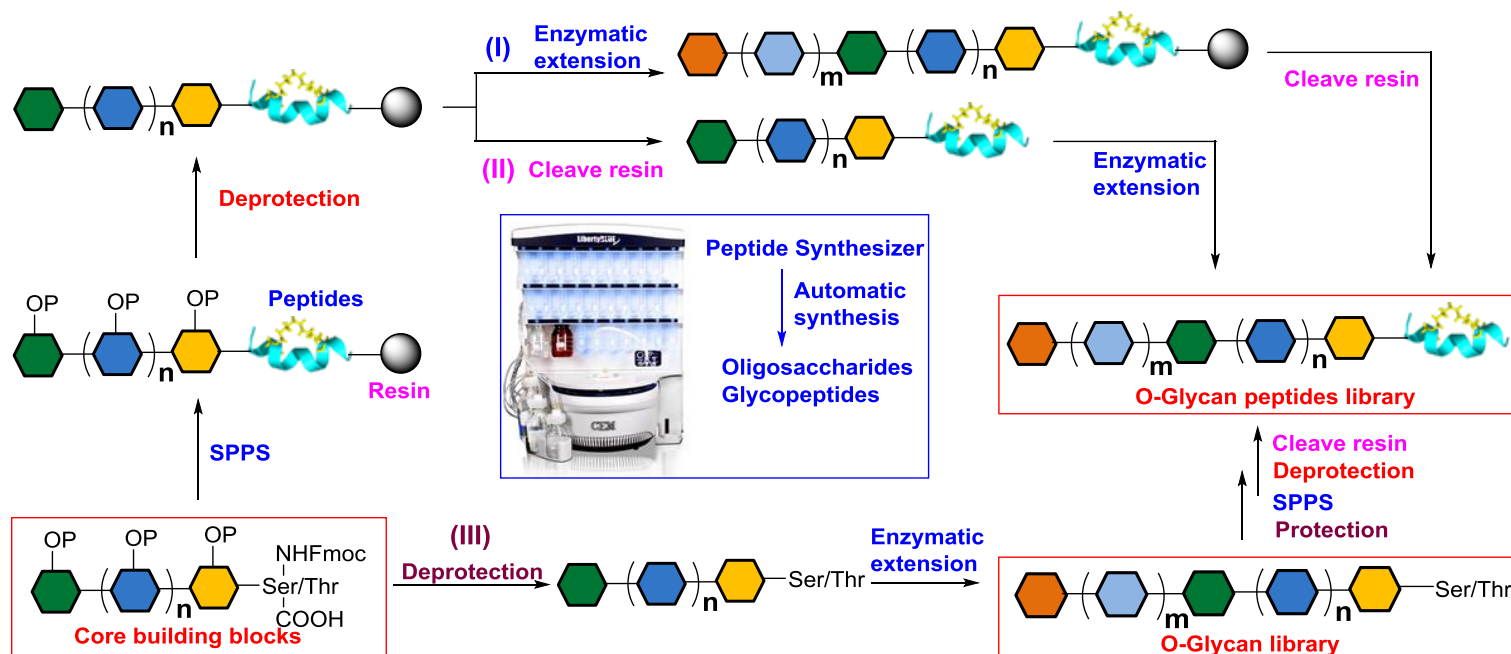


*Enable access of oligosaccharides of defined sequence, branching, and stereochemistry on demand to a diverse range of biomedical researchers.*



# Facile Synthesis of O-glycans & O-glycopeptides

NIH Common Fund Glyco-science Program (U01GM116263)  
Peng George Wang & Lei Li, Georgia State University

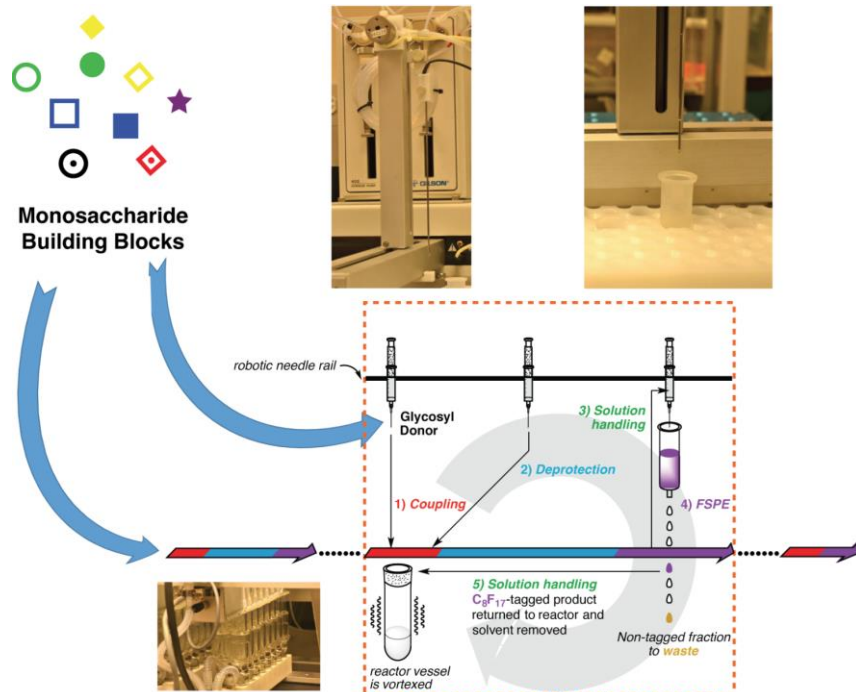


**Goal:** Develop “Core Synthesis/Enzymatic Extension” strategy for the access of O-glycan and O-glycopeptide libraries, automated glycopeptide synthesis

- Convergent chemical synthesis of O-glycan core structures in gram scale;
- Enzymatic extension strategy allows diversity of core structures;
- Automatic glycopeptide synthesis on solid phase/water soluble supports;
- Synthesis of hundreds of O-glycans and O-glycopeptides;
- Cross-validation

# Common Fund Research in the Pohl/Dong Groups: Sugar Building Blocks and Automated Synthesis of Biomedically-Relevant Glycans

*Developing Chemical Methods to Access Building Blocks and Create Oligosaccharides  
Using Solution-Phase Automation Platforms*



*Human Milk Oligosaccharides,  
Bacterial Rhamnans, and  
Mammalian O- and N-Glycans*

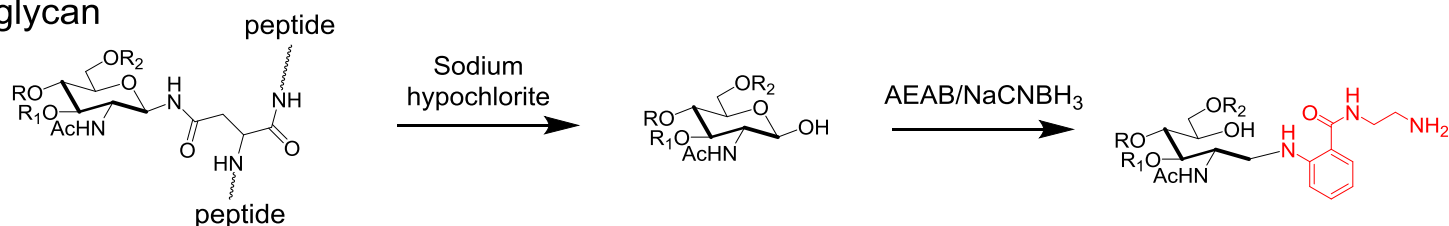
- *Analytical standards and compounds for bioassays with the potential to incorporate fluorescent and other labels*
- *New methods to purify synthetic glycans to 99.5%+ purity for immunological studies (Chem Commun. 2016, 52, 13253)*

<https://commonfund.nih.gov/Glycoscience/fundedresearch#>

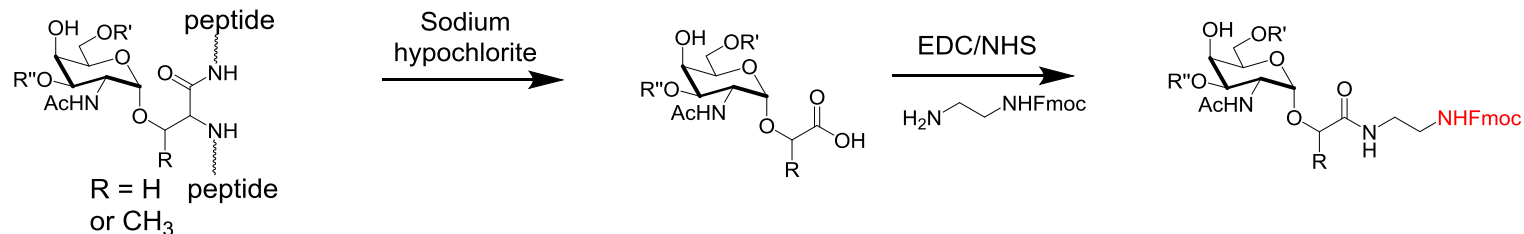


# Large scale chemical preparation of glycans from natural sources

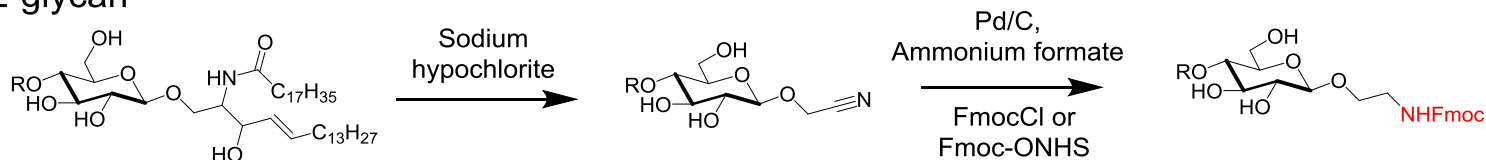
## N-glycan



## O-glycan



## GSL-glycan



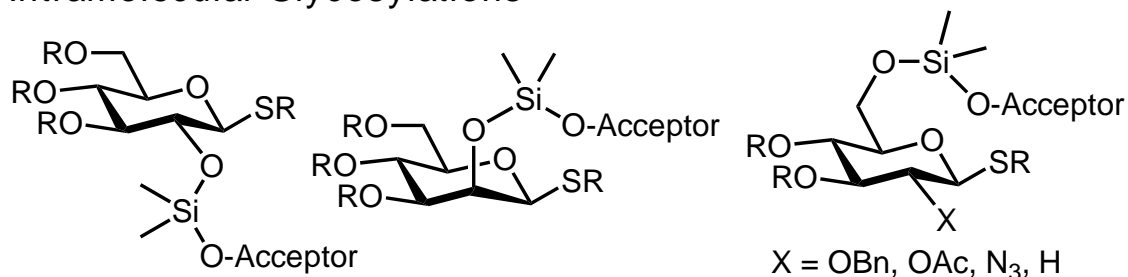
- Novel methods for large scale chemical release and multi-dimensional HPLC separation of natural glycans *Xuezheng Song, Emory University*
- Detailed structure characterization/confirmation of natural glycans *Vernon Reinhold, University of New Hampshire Glycomics Center*



# Catalytic Methods for Building Block Assembly and for Stereoselective Glycosylation (U01GM125274)

John Montgomery and Pavel Nagorny (University of Michigan)

## Intramolecular Glycosylations



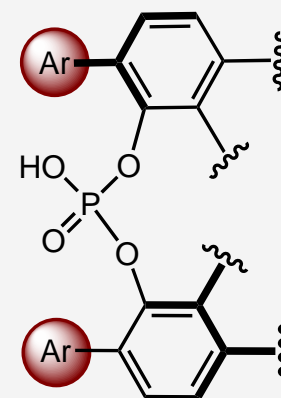
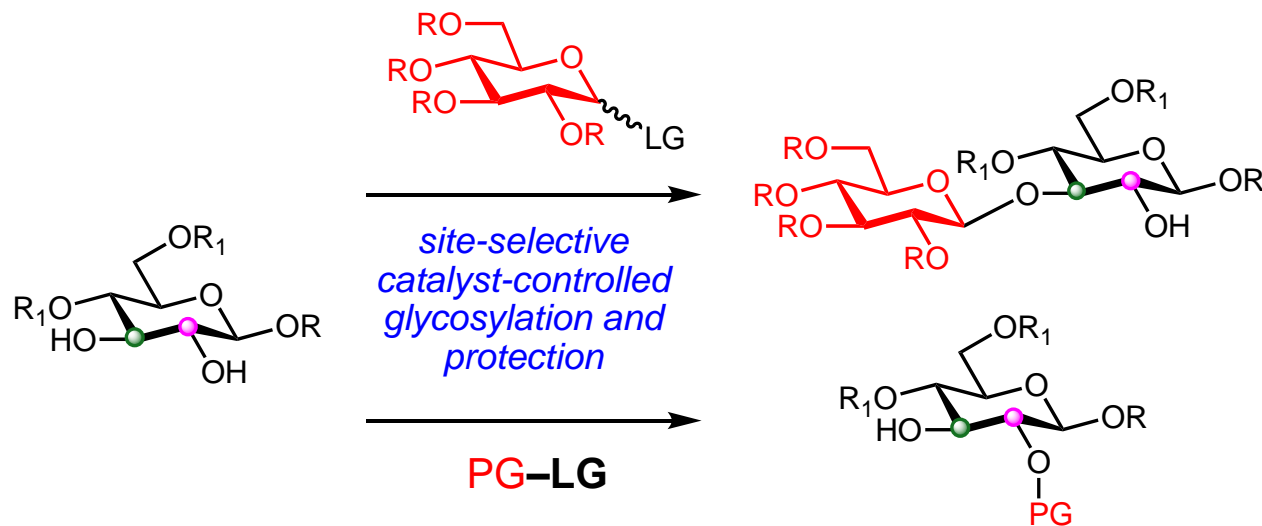
## Accessible Products

Glycoside  
Stereochemistry

b-manno  
a-gluco  
b-gluco

Glycoside  
Functionality

2-benzyloxy  
2-acyloxy  
2-deoxy  
2-azido

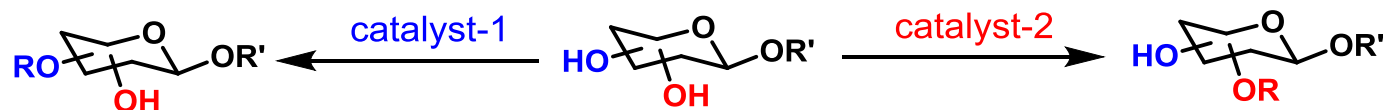


*axially-chiral  
phosphoric  
acids as the catalysts*

## Develop Catalytic Methods to Streamline the Assembly of Oligosaccharides

Weiping Tang, University of Wisconsin – Madison; Peng Liu, University of Pittsburgh

We will develop catalytic site-selective functionalization methods to differentiate hydroxyl groups in various mono- and oligosaccharides in a predictable, general, and systematic manner. The design of these methods will be guided by density functional theory calculations. The resulting transformations will be used for streamlining the synthesis of carbohydrate building blocks.



We will also develop novel transition metal-catalyzed cross-coupling glycosylation (CCG) methods to construct the glycosyl carbon-oxygen bond guided by density functional theory calculations and published literature on cross-coupling reactions. The proposed CCG will allow us to assemble the stereochemically defined benchtop stable glycosyl donors and novel glycosyl acceptors stereospecifically with minimal manipulations after glycosylation.

